Alanya Alaaddin Keykubat University | Rafet Kayış Faculty of Engineering **Genetic and Bioengineering Department** 2023-2024 Fall Semester

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Syllabus						
Code/Name	Sec 403.1 / Cell Culture Techniques					
Type	Elective					
Credit/ECTS	5/5					
Hour per Week	3 (3+0+0)					
Level/Year	Undergraduate/4					
Semester	Fall					
Classroom	D305					
Content	Theoretical and practical introduction to the culture of different types of human and animal cells in the laboratory and how these cells can be transfected to over-express or turn off the expression of different genes.					
Prerequisites						
Textbooks	Primary Culture of Animal Cells: A Manual of Basic Technique, 5th Edition. R. Ian Freshney. Supplementary Cell Culture Manual 3 rd Edition of the SigmaAldrich Cell Culture Manual available free on line: http://www.sigmaaldrich.com/lifescience/cell-culture/learning-center/cell-culture-manual.htm					
Objectives	 The course should provide the student with knowledge such that the student can carry out basic cell-culture techniques properly and safely, and explain factors of significance in the cultivation of cells in vitro. On completion of the course, the student should be able to: - account, at a general level, for the structure and function and maintenance of an LAF/sterile bench Explain how mycoplasma contamination affects eukaryotic cells - account in detail for sterilization equipment and different sterilization techniques Account for different cell culture media and important components in the media Explain the concept of transformation and describe different transformation methods Be able to apply basic cell culture techniques, such as calculation and harvesting of cells 					
Course Outcomes	In this course you will be able to: CO1 describe differences between cell lines and primary cells CO2 describe what is needed to culture cells in the laboratory CO3 describe different types of contaminations that can appear in cell cultures CO4 explain what stem cells are culture cell lines and perform a transfection of cells CO5 choose different ways of culture depending on cell type and purpose CO6 plan a research experiment using mammalian cells, choose a suitable cell system to perform different kinds of experiments.					

Weekly Schedule of Topics

W	Topic			
1	Introduction & biology of cultured cells			
2	Equipments, aseptic techniques, safety protocols			
3	Culture vessels			
4	Media development			

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5	Serum-free medium development			
6	Sterilization			
7	Primary culture			
8	Secondary culture			
9	Cloning & selection			
10	Cell separation, characterization			
11	Differentiation & transformation			
12	Contamination			
13	Cryo-preservation & cyto-toxicity			
14	Organo-typic culture & specialized cell culture techniques			
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Professional	
Contribution	

To have knowledge that will allow working in companies or research laboratories working on cell culture, to be able to design research and projects on cell culture

Contribution to Program Outcomes*

	P01	P02	P03	P04	P05	P06	P07	P08	P09	PO10	P011
CO1	2	2	3	4	2	2	2	3	3	3	4
CO2	2	1	3	3	3	4	3	3	3	0	5
CO3	2	3	2	4	3	4	2	1	2	2	4
CO4	3	3	3	3	3	3	3	2	3	3	4
CO5	3	1	3	4	3	3	1	3	1	2	3
C06	2	2	2	2	3	4	2	2	2	3	2

^{*} Contribution Level | 0: None | 1: Very Low | 2: Low | 3: Medium | 4: High | 5: Very High

Special Conditions	Students work in groups for project and presentations.				
Requirements	Basic knowledge of Cell Biology, Molecular Cell Biology and Biochemistry				
Course Policy	 Be in the class on time. English should always be used to communicate with one another. At least 70% attendance is required, otherwise a grade of DZ will be assigned. 				
Cheating & Plagiarism	 Copying or letting someone copy your work on exams, assignments, or reports is cheating. Cutting and pasting text, figures and tables from web sources or any other electronic source is plagiarism. The consequence of academic dishonesty is to receive a grade of FF for the course. 				
Evaluation	Mid-term Exam 30% Presentation 20% Final Exam 50% Total 100%				
Rubric					

Instructor

Name/Surname	Ayşe Erdoğan	Email	ayse.erdogan@alanya.edu.tr
Room	330	Office Hours	W 13:30-15:30