

Syllabus

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| Code/Name | GBM 302L / GENETIC ENGINEERING II LAB |
| Type | Required |
| Credit/ECTS | 3/3 |
| Hour per Week | 4 |
| Level/Year | Undergraduate/3 |
| Semester | Spring |
| Classroom | FFF L114 |
| Content | This is a course that explains the systems related to the transfer of theoretical knowledge learned in other Genetics and Bioengineering courses into practice. In this context, the necessary materials and processes in laboratory applications will be explained, and students will be able to both design and implement them. The basic plasmid, transfection methodology, cell culture techniques, RNA expression, cell lysis techniques and Western blot study in genetic engineering applications, as well as conventional and modern genetic modification techniques will be explained in detail and with their applications. This course includes basic applications so that students can transfer modern applications of basic experimental design and genetic engineering applications from both academic and industrial perspectives. |
| Prerequisites | GBM 305L GENETIC ENGINEERING I LAB, GBM 305 GENETIC ENGINEERING I |
| Textbooks | <p>Primary Laboratory textbook prepared by the faculty members of our department</p> <p>Supplementary</p> <ul style="list-style-type: none"> • Kurnaz, Isil Aksan. Techniques in Genetic Engineering. 1st ed. CRC Press, 2015. Web. 25 Sept. 2021. ISBN 9780367658816 • Research article published in such library as PUBMED, ELSEVIER |
| Objectives | <ul style="list-style-type: none"> • To teach both the theoretical background and laboratory application principles of the techniques necessary for the processing, modification and artificial transfer of genetic information from cell to cell • To analyze and conduct experimental data • To prepare weekly laboratory report with Genetic and Bioengineering approaches |
| Course Outcomes | <p>In this course students should be able to:</p> <p>CO1. To know laboratory safety rules in Genetic engineering laboratory applications.</p> <p>CO2. To conduct various experiments as cell culture application, transfection, agarose gels, SDS-PAGE gels, cell lysis and protein isolation/amount determination, Western blot studies.</p> <p>CO3. To analyze experimental data, interpret and report experimental results.</p> <p>CO4. To justify the accuracy of experimental results by evaluating them through literature review.</p> |

Weekly Schedule of Topics

| W | Topic | Laboratory Experiments Discussions |
|----|-------|---|
| 1 | | Introduction, organizing study groups, General Information |
| 2 | | Chemical solution preparation techniques |
| 3 | | Introduction to Cell Culture Techniques |
| 4 | | Gene modification (transfection) |
| 5 | | Visualization of PCR products on agarose gel |
| 6 | | Protein isolation types and determination of protein amount |
| 7 | | SDS-PAGE, Western blotting (Only course) |
| 8 | | SDS-PAGE, (Lab) |
| 9 | | Western blotting (Lab) |
| 10 | | Western blotting (Lab) |
| 11 | | Western blotting (Lab) |
| 12 | | Group Presentation |
| 13 | | Group Presentation |
| 14 | | General review |

Professional Contribution Be able to conduct experiment with analyzing data and comparing the results obtained from experiment with literature, and to write weekly report with group member

Contribution to Program Outcomes*

| | PO1 | PO2 | PO3 | PO4 | PO5 | PO6 | PO7 | PO8 | PO9 | PO10 | PO11 |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|------|------|
| CO1 | 0 | 0 | 0 | 0 | 5 | 4 | 2 | 0 | 0 | 0 | 0 |
| CO2 | 3 | 4 | 0 | 3 | 5 | 5 | 3 | 2 | 2 | 0 | 0 |
| CO3 | 3 | 3 | 0 | 4 | 5 | 4 | 4 | 3 | 3 | 0 | 0 |
| CO4 | 2 | 3 | 0 | 4 | 5 | 5 | 5 | 4 | 5 | 2 | 3 |

* Contribution Level | 0: None | 1: Very Low | 2: Low | 3: Medium | 4: High | 5: Very High

Special Conditions

- Students work in groups.
- Experimental studies are reported using MS Word or PDF format.

Requirements

Course Policy

- Students should be in the class or laboratory on time.
- Both student and responsible lecturer should communicate in English
- Students should prepare themselves by reading lab notes and articles sent.
- Students should be prepared by reading the assigned articles and lab notes on a weekly basis.
- At least **80%** attendance is required, otherwise a grade of **DZ** will be assigned.
- Students must be present in class for the presentations, otherwise students will not be graded for the presentation.

Cheating & Plagiarism

- Copying or letting someone copy anyone work on exams, assignments, or reports is cheating.
- Cutting and pasting text, figures and tables from web sources, chatGPT or any other

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Genetic and Bioengineering Department
2023-2024 Spring Semester

electronic source is plagiarism.

- The consequence of academic dishonesty is to receive a grade of **FF** for the course.

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| Evaluation | Laboratory (7×10 pts.) | 8% |
| | Midterm Exam | 32% |
| | Presentation homework | 20% |
| | <u>Final exam</u> | <u>40%</u> |
| | Total | 100% |

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| Rubric | For each report, a rubric will be announced at first week. The rubric has 4 main parts for the grading: aim of the study, material&methods, results and discussion parts |
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| Instructor | | | |
| Name/Surname | Sinem Yilmaz | Email | sinem.yilmaz@alanya.edu.tr |
| Room | 210 | Office Hours | Thursday 10:30-12:30 and 13:30-15:00 |

Prepared by Sinem Yilmaz on June 4th, 2024